

Notification Date: September 5, 2023 Effective Date: October 12, 2023

Mycoplasma genitalium, Transcription-Mediated Amplification, Post-Prostatic Massage Fluid/Urine or Peritoneal Fluids

Test ID: MMGEN

Useful for:

Detecting Mycoplasma genitalium in cases of suspected infection in peritoneal fluid or prostatic secretion (VBIII) fluid/urine

Methods:

Transcription-Mediated Amplification

Reference Values:

Negative

Specimen Requirements:

Submit only 1 of the following specimens:

Specimen Type: Post-prostatic massage fluid/urine (VBIII)

Supplies: Aptima Urine Transport Tube (T582)

Container/Tube: Aptima Urine Specimen Transport Tube

Specimen Volume: 15 to 20 mL

Collection Instructions:

- 1. Patient should not have urinated for at least 1 hour prior to specimen collection.
- 2. Patient should void a small amount of urine prior to prostatic massage. Pre-massage urine can be discarded.
- 3. Patient should cease voiding and a prostatic massage should be performed by urologist or other health care professional.
- 4. Collect post-massage urine into a sterile, plastic, preservative-free container.
- 5. Transfer 2 mL of urine into the Aptima Urine Specimen Transport Tube using the disposable pipette provided within 24 hours of collection. The correct volume of urine has been added when the fluid level is between the black fill lines on the urine transport tube.
- 6. Place the labels on the transport tube so the black fill lines are still visible for volume confirmation at Mayo Clinic Laboratories.
- 7. Transport and store post-prostatic massage fluid/urine (VBIII) specimen transport container at 2 to 30 degrees C (refrigerate is preferred temperature) within 30 days of collection. **Specimen cannot be sent frozen**.

Specimen Type: Peritoneal fluid (pelvic wash, cul-de-sac fluid)

Supplies: Aptima Thin Prep Transport Tube (T652) **Container/Tube:** Aptima Specimen Transfer Tube

Specimen Volume: 1 mL

Collection Instructions:

- 1. Collect specimen by paracentesis using needle and syringe. Refer to the references below.
- 2. Thoroughly vortex specimen upon receipt to the laboratory.

- 3. Transfer 1 mL of specimen into the Aptima Specimen Transfer kit using a disposable transfer pipette or a pipette tip containing a filter (aerosol barrier or hydrophobic plug) within 24 hours of collection.
- 4. Recap the specimen transfer tube tightly.
- 5. Transport and store specimen container at 2 to 30 degrees C (refrigerate is preferred temperature) within 30 days of collection. **Specimen cannot be sent frozen**.

Peritoneal fluid: 1 mL

Post-prostatic massage fluid/urine (VBIII): See Specimen Required

Specimen Stability Information:

Specimen Type	Temperature	Special Container
Varies	Varies	Aptima Vial

Cautions:

This test is intended for non-FDA-approved sources. FDA-approved sources should utilize test AMGEN (Mycoplasma genitalium, Transcription-Mediated Amplification, Varies).

This test does not detect other Mycoplasma or Ureaplasma species.

This test is intended for clinical monitoring or management of patients; it is not intended for use in medico-legal applications.

Results should be interpreted in conjunction with other laboratory and clinical findings.

A negative test result does not exclude the possibility of infection. Improper specimen collection, concurrent antibiotic therapy, presence of inhibitors, or low numbers of organisms in the specimen (ie, below the sensitivity of the test) may cause false-negative test results.

In low prevalence populations, positive results must be interpreted carefully as false-positive results may occur more frequently than true-positive results in this setting.

In general, this assay should not be used to assess therapeutic success or failure, since nucleic acids from these organisms may persist for 3 weeks or more following antimicrobial therapy.

Interference in assay results was observed when mucus at a final concentration of 0.3% weight/volume was added to clinical specimen matrix. Interference was not observed when mucus at a final concentration of 0.03% weight/volume was added to clinical specimen matrix.

Performance of the assay has not been evaluated in individuals younger than 15 years of age.

CPT Code:

87563

Day(s) Performed: Monday through Sunday Report Available: 1 to 4 days

Questions

Contact James Conn, Laboratory Resource Coordinator at 800-533-1710.